POSTER FPN 203P

An Innovative Evidence-Based Laboratory Medicine (EBLM) Test to Help Doctors in Multi-Cancer Early Detection (MCED)

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BACKGROUND

Introduction

- According to the International Agency for Research on Cancer (IARC), **cancer is among the leading causes of death worldwide**[1]. In 2022, there were almost 20 M new cases and 9.7 M cancer-related deaths worldwide. By 2040, new cancer cases per year are expected to rise to 29.9 M and the number of cancer-related deaths to 15.3 M[1]. **Approximately 40.5% of men and women will be diagnosed with cancer at some point during their lifetimes**[1].
- **There are 2 B people worldwide with overweight or obesity —including 124 M children—**, and these figures are expected to continue to increase[2].
- **Obesity is a risk factor for several major cancers**, including post-menopausal breast, colorectal, endometrial, kidney, esophageal, pancreatic, liver, and gallbladder cancer[3]. Excess body fat increases the risk of cancer mortality approximately by 1.75[3].
- **Diet, obesity and lack of physical activity are listed as the main causes of cancer deaths attributed to non-genetic factors**[4]. **The link between obesity and cancer risk is clear**[4].

Key Facts about Cancer in the U.S.

- U.S. cancer endemic: the second leading cause of death[5], and growing.
- In 2024, an estimated 2,001,140 new cases of cancer will be diagnosed in the United States and 611,720 people will die from the disease[5].
- Most common cancers are breast, prostate, lung and bronchus, colon and rectum, melanoma of the skin, bladder, kidney and renal pelvis, non–Hodgkin lymphoma, endometrial, pancreatic, leukemia, thyroid, and liver cancer[5].

Most Studied Known or Suspected Cancer Risk Factors[6]

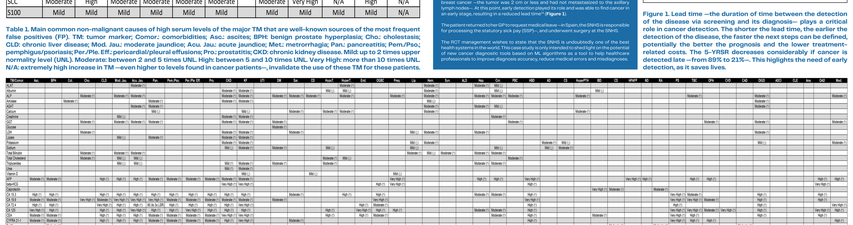
- Age
- Alcohol
- Cancer-causing substances
- Chronic inflammation
- Diet
- Hormones
- Immunosuppression
- Infectious agents
- Overweight and obesity
- Radiation
- Sunlight
- Tobacco

Cancer Prognosis and 5–Year Relative Survival Rate (5–YRSR)[7]

For all types of cancer, **the 5-YRSR is strongly associated with tumor stage at detection**.

In this way, the 5-YRSR for localized cancers —detected early, when cancer is most likely to be treated with success— is the highest —up to the 99.9% for papillary thyroid carcinoma (PTC)—. However, **when cancer has metastasized to other parts of the body, the 5-YRSR drops** —to below 3.1% for metastasic lung cancer (LC)—. Apart from the stage of the cancer at the time of detection, many other factors such as the patient's age and health can also significantly affect the 5-YRSR.

The 5-YRSR for some of the most frequent solid tumors, according to their stage at diagnosis —localized, regional and distant—, are[7] (alphabetically sorted):



Current Tools to Diagnose Cancer

They are often invasive, expensive and may give many false positives (FP):

- TM laboratory tests (blood, urine and other fluid samples)[8,10,11]: cheap and fast, but inaccurate, as **other comorbidities, apart from cancer, can also increase serum TM levels**, thus **giving FP and reducing the specificity**.
- X-Ray: **very low accuracy** —high rate of FP due to scars, cysts or benign nodules—, and **radiation exposure**.
- Algorithms developed by the American College of Radiology (ACR)[12] for imaging procedures: BI-RADS for mammography, ultrasound (US) or magnetic resonance imaging (MRI); C-RADS[13] for computerized tomography (CT) scan; GI-RADS for transvaginal (TV) US; LI-RADS® for US or CT-scan; Lung-RADS for CT-scan; O-RADS™ for US; PI-RADS® for MRI; or TI-RADS™ for ultrasonography. They are usually **expensive** and they have a **high FP rate**, which may lead to unnecessary invasive biopsies.
- Liquid biopsy: blood test that looks for circulating tumor DNA (ctDNA). Its **accuracy can vary depending on the type and stage of cancer, detecting low levels of ctDNA in early-stage cancers. Very expensive**.
- Tissue biopsy: **current gold standard in cancer diagnostics**, offering a detailed view of the tumor's cellular makeup. However, they are **highly invasive and expensive**.

Upcoming Non–Invasive Cancer Diagnosis Solutions

In recent years, tumor markers (TM) have been suggested to help in cancer recurrence surveillance, confirmation diagnosis and screening.

However, **TM can be also increased due to non-malignant diseases**[8,11] **(Table 1), mainly in the liver and kidneys, since TM are mainly catabolized in the liver —detoxification pathway—, and excreted through the kidneys**.

Therefore, any disease that affects these two organs will affect TM serum levels[8,11]. In this way, the **Spanish Society of Laboratory Medicine (SEQC^{ML})** developed in 1994 a set of rules known as the **Barcelona Criteria**[9] to minimize FP:

1. Evaluate **serum TM concentration**.
2. Exclude the main source of FP results by **discarding benign pathologies (Table 1)**.
3. When moderate results, **perform follow-up —serial testing—**.
4. Eliminate high results by **avoiding technical interferences**.

TM/Comor.	Asc.	BPH	Cho.	CLD	Mod. Jau.	Acu. Jau.	Met.	Pan.	Pem./Pso.	Per./Pe. Eff.	Pro.	CKD
AFP	Moderate	Moderate		High	High	High			Moderate	Moderate	Moderate	Moderate
CA 15.3	High	High	High	High	High	High	High	High	High	High	High	High
CA 19.9	Moderate	Moderate	Very High	Moderate	Very High	Very High	Very High	Very High	Moderate	Moderate	Moderate	Moderate
CA 72.4	High	High		Very High	High	High			High	High	High	High
CA 125	Very High	High		High	High	Very High	High		High	High	High	High
CEA	Moderate	Moderate		Moderate	Moderate	Moderate			Moderate	Moderate	Moderate	Very High
CYFRA 21-1	Moderate	Moderate		High	High	High			Moderate	Moderate	Moderate	Moderate
HE4	Moderate	Mild		High	High	High			Mild	Mild	Mild	N/A
NSE	Moderate	Moderate	Moderate	Moderate	Moderate	Moderate			Moderate	Moderate	Moderate	Moderate
ProGRP	High	High		High	High	High			High	High	High	Very High
PSA	Moderate	Very High	Moderate	Moderate	Moderate	Moderate			Moderate	Very High	Moderate	Moderate
SCC	Moderate	High		Moderate	Moderate	Moderate			Moderate	Very High	N/A	High
S100	Mild	Mild		Mild	Mild	Mild			Mild	Mild	Mild	N/A

Table 1. Main common non-malignant causes of high serum levels of the major TM that are well-known sources of the most frequent false positives (FP). TM: tumor marker; Comor: comorbidities; Asc.: ascites; BPH: benign prostatic hyperplasia; Cho.: cholecystitis; CLD: chronic liver disease; Mod. Jau.: moderate jaundice; Acu. Jau.: acute jaundice; Met.: metrorrhagia; Pan.: pancreatitis; Pem./Pso.: pemphigus/pouretic fever; Per./Pe. Eff.: pericardial / pleural effusions; Pro.: prostatitis; CKD: chronic kidney disease. Mild: up to 2 times upper normality level (UNL). Moderate: between 2 and 5 times UNL. High: between 5 and 10 times UNL. Very High: more than 10 times UNL. N/A: extremely high increase in TM —very higher to levels found in cancer patients—, invalidate the use of these TM for these patients.

Case Study

A 51 y.o. women got tested and the algorithm suggested that a cancer was present, with a 97% probability, but without establishing the site. Nonetheless, it suggested that it could be a breast or a digestive cancer. Besides, the algorithm also suggested hepatic disease, specifically due to alcohol abuse —even though the patient stated "I swallowed a sip of alcohol when I was 20 years old"—. The algorithm is as a tool to help healthcare professionals to improve diagnosis accuracy, reduce medical errors and misdiagnoses, as well as avoiding invasive procedures.

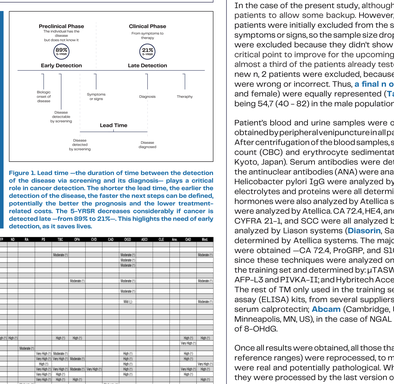


Figure 1. Lead time —the duration of time between the detection of the disease via screening and its diagnosis— plays a critical role in cancer detection. The shorter the lead time, the earlier the detection of the disease, the faster the next steps can be defined, potentially the better the prognosis and the lower treatment-related costs. The HBP disease example: Currently if cancer is detected late —from 69% to 21%—. This highlights the need of early detection, as it saves lives.

METHODS

This study builds upon the knowledge gained from the prior development of the algorithm for the detection of cancer of unknown primary (CUP) —a non-invasive blood and urine test to help in differential diagnosis of patients with one or more metastatic sites, but without the primary site identified—, and by adding other specific algorithms for the screening of different types of cancer. The latter have been presented and published at several prestigious scientific congresses, such as: **American Society of Clinical Oncology (ASCO)** 2022 (breast[21], colon[22]), European Society for Medical Oncology (ESMO) Gastrointestinal Cancers Congress 2024 (liver[23]), ESMO Gynecological Cancers Congress 2024 (ovary[24]), and International Association for the Study of Lung Cancer (IASLC) World Conference on Lung Cancer (WCLC) 2024 (lung[25]). Besides, it is also based on different clinical research studies carried out over the last eight years. Moreover, it is also based on the **Barcelona Criteria**, in order to improve its screening accuracy by avoiding FP[20–28].

Even though in the algorithm modeling set, several innovative and/or experimental TM —serum calprotectin, alfa-fetoprotein L3 fraction (AFP-L3), cystein-rich angiogenic inducer 61 (Cyr61), epidermal growth factor receptor (EGFR), human kallikrein 2 (HK2), neutrophil gelatinase–associated lipocalin (NGAL), protein induced by vitamin K absence or antagonist–II (PIVKA–II), [–2]proPSA (p2PSA), soluble Axl (sAxl), and 8-Hydroxi–2'–deoxyguanosine (8–OHdG)—, were included, this study was developed only with the 15 most routinely used TM — alfa-fetoprotein (AFP), beta human chorionic gonadotropin (beta-hCG), carbohydrate antigen 15.3 (CA 15.3), carbohydrate antigen 19.9 (CA 19.9), carbohydrate antigen 72.4 (CA 72.4), carbohydrate antigen 125 (CA 125), carcinoembryonic antigen (CEA), cytokeratin 19 fragment 21–1 (CYFRA 21–1), human epidydymis 4 (HE4), neuron-specific enolase (NSE), pro–gastrin–releasing peptide (ProGRP), prostate specific antigen total (PSA total), prostate specific antigen free (PSA free), squamous cell carcinoma (SCC) antigen, and S100 protein (S100). Besides, the whole panel is based on many different specific panels which, in turn, include many other clinical analytes —**to meet the Barcelona Criteria in order to minimize FP**—, such as: hematology, antibodies, electrolytes, proteins, hepatic enzymes, hormones, vitamins, and other specific serum and urine analytes, as well as urinalysis.

To develop the algorithm, all data was computed firstly with the **Multi-Biomarker Disease Activity Algorithm (MBDAA)**[29] —developed by **Bioprognos** (Barcelona, Spain)—. Besides, a couple of new machine learning algorithms (ML) were also included, such as **Evidence-Based Laboratory Medicine Algorithm (EBLMA)**[30] and **Artificial Intelligence Recursive Algorithm (AIRA)**[31] —both developed by **Blueberry Diagnostics** (Barcelona, Spain)—, to improve both sensitivity and specificity. EBLMA is the algorithm that includes the **Barcelona Criteria** basis already upgraded with more knowledge regarding the correct interpretation of TM and the elimination of FP. EBLMA include several functions —sub-algorithms— to detect the following well-known conditions: iron transport and storage function diseases —iron deficiency and anemia (distinguishing between microcytic, normocytic, and macrocytic); cardiovascular disease (CVD) and atherosclerotic cardiovascular disease (ASCVD); pancreatic endocrine diseases —insulin resistance (IR), prediabetes, diabetes mellitus (DM)—; thyroid function diseases —primary, secondary and tertiary hypo and hyperthyroidism, both of non-autoimmune or autoimmune etiology—; parathyroid function and bone metabolism diseases —hyposparathyroidism and primary or secondary hyperparathyroidism, as well as osteopenia and osteoporosis (Ops)—; vitamin metabolism diseases —vitamin B12 deficiency and insufficiency, and vitamin D deficiency and insufficiency—; hepatic function diseases —non-alcoholic fatty liver disease (NAFLD), non-alcoholic steatohepatitis (NASH), liver fibrosis and cirrhosis—; renal function diseases —chronic kidney disease (CKD)—; electrolyte imbalance–related diseases —hypo or hypercalcemia, hypo or hypercholesterolemia, hypo or hypermagnesemia, hypo or hypernatremia, hypo or hyperphosphatemia, hypo or hyperpotassemia, or hypo or hyperuricemia—; prostate diseases —benign prostatic hyperplasia (BPH), prostatitis and prostatic intraepithelial neoplasia (PIN)—; ovarian diseases —endometriosis and polycystic ovary syndrome (PCOS)—; and inflammatory response diseases —rheumatoid arthritis—. In the other hand, AIRA is an algorithm that computes in a recursive way all the values from 0 to 100% for both sensitivity and specificity to find the optimal cutoff values for low, moderate (moderate–low and moderate–high), and high-risk results. After that, **parallel approximations to optimize overall sensitivity (Se) were conducted**. Finally, **serial approximations to enhance specificity (Sp) were performed** —both powered by a tailor–made ML algorithm—.

Following the above mentioned process, we were able to determine an initial sensitivity and specificity of the algorithm with a training set of 192,090 patients. This set was very heterogenous, as it included results analyzed by different suppliers for the same laboratory determinations —Roche, Siemens Healthineers, Beckmann or Abbott, among others—, with their own systems and reference values. So, the variability and the potential biases in the results obtained by this way may be too high, thus affecting the quality of the results. In this way, to avoid these potential biases that can alter the results —the accuracy of the algorithm—, the present study was designed with one single laboratory —**Laboratorio Echevarne** (Sant Cugat del Vallès, Barcelona, Spain)—, and consisted in a randomized controlled trial (RCT) with a target sample size (n) of 300.

All patients included in the RCT were recruited through 3 private medical centres in Barcelona (Spain). Patients had to accept and sign the informed consent, the ethics comittee approvement, and the test requisition form (TRF) —an example of the new TRF designed for the upcoming RCT that will be performed in the U.S. with **Empire City Laboratories** (Brooklyn, NY, US) is shown (**Figure 3**)—.

In the case of the present study, although target sample size was 300 patients, we managed to enroll 314 patients to allow some backup. However, according to the inclusion and exclusion criteria (**Figure 2**), 36 patients were initially excluded from the study, because they had ongoing clinically diagnosed pathologies, symptoms or signs, so the sample size dropped to 278 patients. From this new study population, 110 patients were excluded because they didn't show up at the clinical facility for any of the follow-up visits —this is a critical point to improve for the upcoming RCT, since the cost of each patient is very high and for this RCT almost a third of the patients already tested were lost—, so the new n consisted of 168 patients. From this new n, 2 patients were excluded, because some of their laboratory parameters and/or clinical information were wrong or missed. In this way, a total of 166 patients (mean age = 60 years; range = 18 to 85 years; male and female) were equally represented (**Tables 3–9**) and the mean age of the participants was 53,1 years, being 54,7 (40 – 82) in the male population and 51,5 (40 – 77) in the female population (**Tables 3, 5**).

Patient's blood and urine samples were obtained from October 2021 to June 2023. Blood samples were obtained by peripheral venipuncture in all participants. All analytics were performed in **Laboratorio Echevarne**. After centrifugation, serum and plasma were separated from whole blood. Erythrocyte sedimentation rate (serum cube) (CBC) and erythrocyte sedimentation rate (ESR) were determined by Yumizen analyzers (**Horiba**, Kyoto, Japan). Serum antibodies were determined by Atellica analyzers (**Siemens**, Munich, Germany), but the antinuclear antibodies (ANA) were analyzed by Elecsys systems (**Roche**, Rotkreuz, Switzerland) and the Helicobacter pylori IgG was analyzed by Vidas systems (**bioMérieux**, Marcy-l'Étoile, France). The serum electrolytes and proteins were all determined by the Atellica analyzers. All the serum hepatic enzymes and hormones were also analyzed by Atellica systems. In the case of TM, AFP, beta-hCG, PSA total, and PSA free were analyzed by Atellica systems. The rest of TM —CA 72.4, ProGRP, and S100— were analyzed by Elecsys 8,3, CA 15.3, CYFRA 21–1, and SCC were all analyzed by Alinity systems (**Abbott**, Chicago, IL, US). NSE and S100 were analyzed by Liaison systems (**Diasorin**, Saluggia, Italy). The remaining serum and urine analytes were also determined by Atellica systems. The majority of the tests were performed within one hour after samples were obtained —CA 72.4, ProGRP, and S100 were performed within a week after samples were obtained since these techniques were analyzed once a week—. On the other hand, novel TM were only analyzed in the training set and determined by: µTASWako i30 (**Fujifilm Medical Systems**, Tokyo, Japan), in the case of AFP–L3; ImmuTOP i-WAK-IT, and Wyntech Access (**Beckman Coulter**, Brea, CA, US), in the case of HK2 and p2PSA. The rest of TM only used in the training set were analyzed by commercial enzyme-linked immunosorbent assay (ELISA) kits, from several suppliers, such as: **Bühlmann** (Schönenbuch, Switzerland), in the case of serum calprotectin; **Cayman Chemical** (Ann Arbor, MI, US), in the case of Cyr61; **Fine Test** (Wuhan, China), in the case of EGFR; **Abnova** (Taipei, China), in the case of HK2; **Bosche**, Minneapolis, MN, US), in the case of NGAL and sAxl; and **Cayman Chemical** (Ann Arbor, MI, US), in the case of 8–OHdG.

Once all results were obtained, all those that exhibited abnormal values (that is results that were outside their reference ranges) were reprocessed, to make sure that they were not obtained due to technical errors and were real and potentially pathological. When all laboratory determinations were finished per each patient, they were processed by the last version of the algorithm during the final results.

Several calculations —derived, mainly, from values related with other ratios or indices (**Table 6**), hepatic and biliary tract indices (**Table 4**), pancreatic endocrine indices (**Table 7**), and renal indices (**Table 8**). Besides, a few inflammatory ratios (**Table 9**) were also performed —many studies have suggested their use as a novel tool in the assessment of many malignancies—. We selected the ones that were **Evidence-Based Laboratory Medicine (EBLM)** and **Artificial Intelligence (AI)**–driven. In this way, pancreatic endocrine disease criterion (AC)[32,43], atherogenic index of plasma (AIP)[33–45], Castelli risk index I[46,47], Castelli risk index I + bilirubin[47], Castelli risk index II[47], Castelli risk index II + bilirubin[47], cholesterol/bilirubin ratio[47], lipoprotein combine index (LPCI)[48], metabolic syndrome (MetS)[49], LTI[49,50], and triglycerides/HDL–c index[51]. Besides, pancreatic endocrine indices performed were adult treatment panel III (ATP–III)[52,53] —metabolic syndrome—, triglycerides and glucose (TyG) index[54–56] —insulin resistance—, homeostatic model assessment (HOMA–IR)[57,58], homeostatic model assessment 2 (HOMA–2)[59], and quantitative insulin sensitivity check index (QUICKI)[60]. Besides, hepatic and biliary tract indices performed were: AST/ALT ratio[61], de Ritis ratio[62], hepatic steatosis index (HSI)[63,64], K–NALFD score[65], NAFLD liver fat score (LFS)[66], NAFLD logit score (NLS)[65], NAFLD ridge score (NRS)[67], and ALD NAFLD index (ANI)[67] for the assessment of non-alcoholic fatty liver disease (NAFLD); fibrosis–4 index (FIB–4)[68], King's score[69], Lok index[70], and Forns index[71] for the assessment of hypertension–ALT–insulin resistance (HAIR) score[66] for the assessment of non-alcoholic steatohepatitis (NASH); AST–to–platelet ratio index (APRI)[72,73], BMI–age–ALT–triglycerides (BAAT) score[66], BMI–AST/ALT ratio–diabetes (BARD) score[74], ELF test[75], FAST score[76], FIB 4 score[77], Hepamet fibrosis score[78], nonalcoholic fatty liver disease (NFS)[69,79], and steatosis–associated fibrosis estimator (SAFE) score[80] for the assessment of liver fibrosis and cirrhosis, R Factor[73], and chronic liver disease (CLivD) score[81] were also performed for further risk–assessment. Furthermore, inflammatory ratios performed were: RDW–to–LYM[82], RDW–to–NEU[82], RDW–to–MON[82], RDW–to–RBC[82], RDW–to–PLT[77], RDW–to–PDW[77], Hb–to–RDW[83], LYM–to–MON[82], MON–to–LYM[82], MON–to–NEU[77], MPV–to–LYM[85], MPV–to–NEU[77], MPV–to–MON[77], MPV–to–PLT[77], NEU–to–LYM[77,82], Derived NEU–to–LYM[77,83], NEU–to–LYM+PLT[84], NEU–to–MON[77,82], PDW–to–LYM[85], PDW–to–NEU[77], PDW–to–MON[77], PLT–to–WBC[77], PLR–to–NEU[86], PLR–to–MON[86], PLR–to–LYM[86], ALP–to–RBC[77], ALP–to–NEU[77], ALP–to–MON[77], ALP–to–LYM[77], ALP–to–PLT[77,82], ALP–to–PDW[77], LDH–to–RBC[77], LDH–to–RDW[77,83], LDH–to–LYM[77,83], LDH–to–PLT[77,83], LDH–to–PDW[77], LDH–to–albumin[87], NEU–to–HDL–c[84], MON–to–HDL–c[84,88], PLT–to–HDL–c[88], prognostic nutritional index (PNI)[89], CA 15.3–to–RBC[77], CA 15.3–to–PLT[77], and CA 15.3–to–PDW[77]. Finally, renal indices performed were: corrected electrolytes[90–92], creatinine clearance[93], eGFR[94], BUN–to–creatinine ratio[93], and urine albumin–to–creatinine ratio (ACR)[94]. Then, the cancer–related functions of the algorithms were executed.

In turn, all patients with suspicious results were referred to the corresponding medical centres for confirmation and subsequent classification in each of the groups —case and control—, designed for the biostatistics upcoming phase. Patients attended follow–up visits in one year.

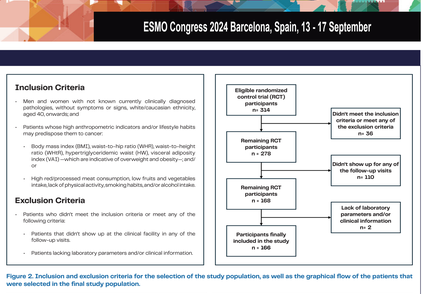


Figure 2. Inclusion and exclusion criteria for the selection of the study population, as well as the graphical flow of the patients that were selected in the final study population.

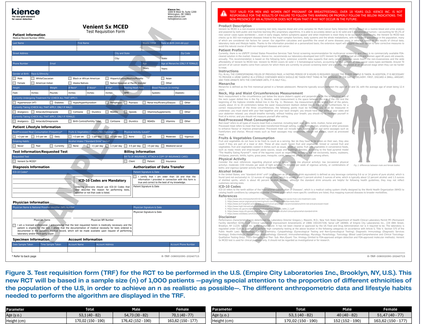


Figure 3. Test requisition form (TRF) for the RCT to be performed in the U.S. (Empire City Laboratories Inc., Brooklyn, NY, U.S.). This new RCT will be based in a sample size (n) of 1,000 patients —paying special attention to the proportion of different ethnicities of the population of the U.S. in order to achieve an n as realistic as possible—. The different anthropometric data and lifestyle habits needed to perform the algorithm are displayed in the TRF.

Parameter	Total	Male	Female
Age (y.o.)	53,1 (40 – 82)	54,7 (40 – 82)	51,5 (40 – 77)
Height (cm)	170,1 (150 – 195)	174,2 (157 – 195)	165,6 (150 – 180)
Weight (kg)	71,36 (38,6 – 128)	82,90 (56,6 – 128)	59,77 (40 – 112)
Neck Circumference (cm)	36,49 (29 – 50)	40,3 (34 – 47)	32,76 (29 – 50)
Waist Circumference (cm)	86,34 (62,5 – 131)	96,77 (66,5 – 131)	76,60 (62,5 – 131)
Hip Circumference (cm)	101 (76 – 131)	104,5 (86 – 131)	96,36 (76 – 131)
Hemanthe (y.o.)	NA	NA	13,68 (9 – 17)
Body Mass Index (BMI)	117,7 (71,1 – 176)	117,7 (71,1 – 176)	117,7 (71,1 – 176)
Diastolic Pressure (mmHg)	70,62 (50 – 100)	73,29 (50 – 100)	76,76 (53 – 100)
Resting Heart Rate (BPM)	69,33 (42 – 110)	66,76 (42 – 110)	69,75 (50 – 99)
Waist–to–Hip Ratio (WHR)	0,85 (0,74 – 0,97)	0,92 (0,74 – 0,97)	0,79 (0,74 – 0,84)
Hemoglobin (g/dL)	13,74 (7,3 – 17,3)	14,9 (7,7 – 17,3)	12,68 (9,3 – 16,8)
Hematocrit (%)	41,1 (23,1 – 51,4)	44,3 (23,1 – 51,4)	37,9 (28,1 – 47,6)
Erythrocytes (M/mm3)	4,66 (2,62 – 6,06)	5,01 (2,62 – 6,06)	4,31 (3,19 – 5,42)
ESR (mm)	8,43 (2 – 39)	6,83 (2 – 33)	9,80 (2 – 39)
MCV (fL)	88,50 (76,2 – 97,7)	88,44 (76,2 – 97,7)	88,56 (77,1 – 97,6)
MCH (pg)	29,57 (24,8 – 33,6)	29,74 (24,8 – 33,6)	29,39 (24,8 – 33,6)
MCHC (g/L)	334,1 (316,4 – 348,6)	336,2 (316,4 – 348,6)	331,7 (316,4 – 348,6)
Leukocytes (K/mm3)	6,42 (2 – 39)	6,83 (2 – 33)	5,80 (2 – 39)
Neutrophils Absolute (x10^3/µL)	3,77 (1,51 – 11,94)	3,96 (1,96 – 11,94)	3,55 (1,51 – 9,75)
Neutrophils Relative (x10^3/µL)	3,84 (1,51 – 9,94)	3,93 (1,96 – 9,94)	3,72 (1,51 – 9,75)
Lymphocytes Absolute (x10^3/µL)	1,53 (0,31 – 2,69)	1,7 (0,76 – 2,75)	1,34 (0,31 – 2,69)
Lymphocytes Relative (x10^3/µL)	1,53 (0,31 – 2,69)	1,7 (0,76 – 2,75)	1,34 (0,31 – 2,69)
Monocytes Absolute (x10^3/µL)	0,17 (0,01 – 0,91)	0,2 (0,02 – 0,91)	0,13 (0,01 – 0,91)
Monocytes Relative (x10^3/µL)	0,17 (0,01 – 0,91)	0,2 (0,02 – 0,91)	0,13 (0,01 – 0,91)
Eosinophils Absolute (x10^3/µL)	0,09 (0,01 – 0,46)	0,11 (0,01 – 0,46)	0,07 (0,01 – 0,46)
Eosinophils Relative (x10^3/µL)	0,09 (0,01 – 0,46)	0,11 (0,01 – 0,46)	0,07 (0,01 – 0,46)
Basophils Absolute (x10^3/µL)	0,04 (0,01 – 0,2)	0,04 (0,01 – 0,2)	0,04 (0,01 – 0,2)
Basophils Relative (x10^3/µL)	0,04 (0,01 – 0,2)	0,04 (0,01 – 0,2)	0,04 (0,01 – 0,2)
Platelet Count (x10^3/µL)	259,47 (136 – 442)	238,36 (136 – 442)	286,93 (144 – 570)
Platelet Volume (fL)	9,45 (6,7 – 14)	9,45 (6,7 – 14)	9,45 (6,7 – 14)
Plateletcrit (%)	0,24 (0,09 – 0,42)	0,23 (0,09 – 0,42)	0,27 (0,09 – 0,42)
Platelet Distribution Width (PDW)	16,39 (9,2 – 18,4)	16,39 (9,2 – 18,4)	16,39 (9,2 – 18,4)
Platelet–Large Cell Ratio (P–LCR)	34,85 (7,6 – 56)	34,85 (7,6 – 56)	34,85 (7,6 – 56)
Platelet–to–Lymphocyte Ratio (PLR)	169,51 (61,9 – 1.042)	139,61 (61,9 – 1.042)	203,41 (74,1 – 1.042)
Lymphocyte–to–Monocyte Ratio (LMR)	35,26 (6,3 – 94)	39,68 (6,8 – 94)	30,83 (11,2 – 66,2)
Neutrophils Ratio (%)	5,75 (2 – 12,6)	6,27 (4,4 – 11,3)	5,27 (2 – 12,6)
Lymphocytes (%)	36,14 (11 – 60)	33,1 (14 – 60)	39,18 (20 – 59)
Monocytes (%)	3,97 (0,4 – 12,3)	3,93 (0,4 – 12,3)	4,01 (0,4 – 11,2)
Eosinophils (%)	2,13 (0,1 – 10,4)	2,17 (0,1 – 10,4)	2,09 (0,1 – 10,4)
Basophils (%)	0,97 (0,1 – 2,3)	0,93 (0,1 – 2,3)	1,01 (0,1 – 2,3)
Urine Color	Yellow	Yellow	Yellow
Urine Aspect	Clear	Clear	Clear
Urine Density	1,02 (1,01 – 1,03)	1,02 (1,01 – 1,03)	1,02 (1,01 – 1,03)
Urine pH	5,77 (5 – 7,5)	5,86 (5 – 7,5)	5,69 (5 – 7,5)
Urine Nitrites	0,04 (0 – 1)	0,03 (0 – 1)	0,06 (0 – 1)
Urine Leukocytes	0,07 (0 – 1)	0,06 (0 – 1)	0,08 (0 – 1)
Urine Glucose	0,01 (0 – 1)	0,01 (0 – 1)	0,01 (0 – 1)
Urine Ketones	0,07 (0 – 1)	0,06 (0 – 1)	0,07 (0 – 1)
Urine Proteins	0,04 (0 – 1)	0,05 (0 – 1)	0,03 (0 – 1)
Urine Urobilin	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Urine Bilirubin	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)

Table 3. Raw data —clinical and laboratory data—, obtained, displayed as the mean value of each parameter with the corresponding minimal (Min) and maximal (Max) value, for all patients and by gender: male and female.

Parameter	Total	Male	Female
Albumin/Globulin Ratio (A/G)	1,51 (0,7 – 2,4)	1,49 (0,7 – 2,4)	1,53 (0,8 – 2,4)
Albumin/Globulin Ratio (A/G) Abnormal	0,01 (0 – 1)	0,01 (0 – 1)	0,01 (0 – 1)
Albumin/Globulin Ratio (A/G) Normal	0,99 (0 – 1)	0,99 (0 – 1)	0,99 (0 – 1)
Albumin/Globulin Ratio (A/G) Status	0,99 (0 – 1)	0,99 (0 – 1)	0,99 (0 – 1)
Albumin/Globulin Ratio (A/G) Status Abnormal	0,01 (0 – 1)	0,01 (0 – 1)	0,01 (0 – 1)
Albumin/Globulin Ratio (A/G) Status Normal	0,99 (0 – 1)	0,99 (0 – 1)	0,99 (0 – 1)
Albumin/Globulin Ratio (A/G) Status Undetermined	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Status	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Status Abnormal	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Status Normal	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Status Undetermined	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Undetermined	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Undetermined Status	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Undetermined Status Abnormal	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Undetermined Status Normal	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Undetermined Status Undetermined	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Undetermined Undetermined	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Undetermined Undetermined Status	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Undetermined Undetermined Status Abnormal	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Undetermined Undetermined Status Normal	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Undetermined Undetermined Status Undetermined	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Undetermined Undetermined Undetermined	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Undetermined Undetermined Undetermined Status	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Undetermined Undetermined Undetermined Status Abnormal	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G) Undetermined Undetermined Undetermined Undetermined Status Normal	0,0 (0 – 0)	0,0 (0 – 0)	0,0 (0 – 0)
Albumin/Globulin Ratio (A/G)